

New Regulatory Opportunities in DART

For Pharmaceutical Development

GOOD
MEDICINES
USED
BETTER

A Short History of DART testing

Thalidomide (softenon in NL)

- Late 1950's on the market as drug against morning sickness
- Caused impaired limb growth in human embryo's (Phocomelia)
- Increased awareness of possible effects of drugs and chemicals on embryonic development

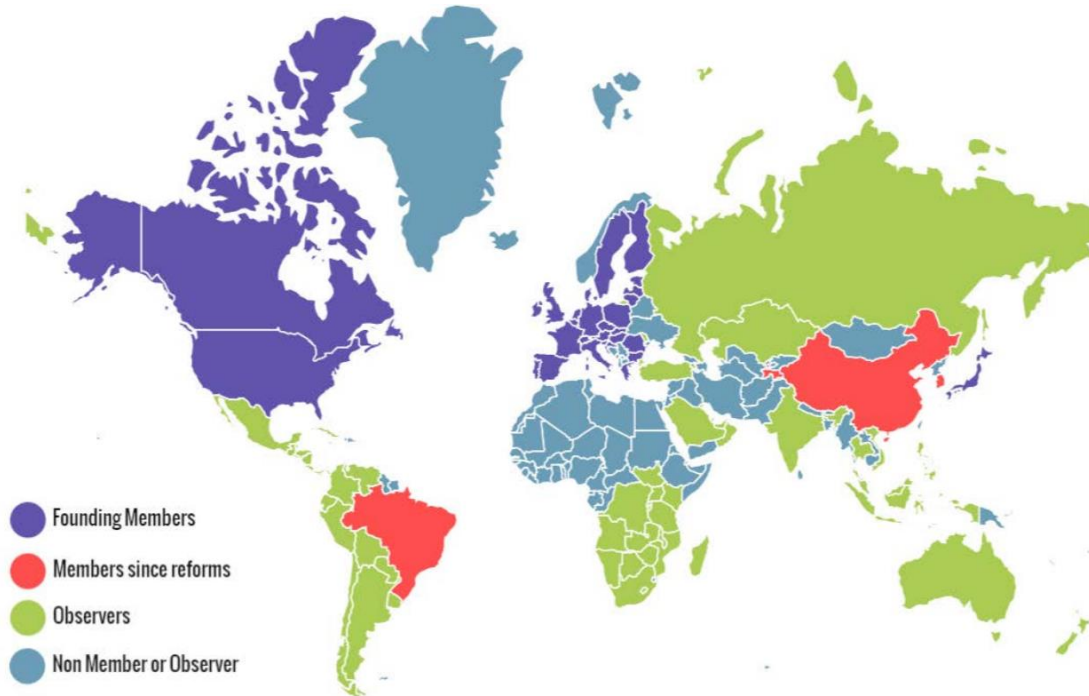
Thalidomide tragedy led to safety guidance for DART testing:

→ 1970s first guidelines for reproductive toxicity testing for medicines (FDA)

→ in 1980s for chemicals (OECD)

→ Global Harmonized DART Guideline for Pharmaceuticals in 1995 (ICH S5)





INTERNATIONAL COUNCIL FOR HARMONISATION OF TECHNICAL
REQUIREMENTS FOR PHARMACEUTICALS FOR HUMAN USE

ICH HARMONISED GUIDELINE

**DETECTION OF REPRODUCTIVE AND DEVELOPMENTAL
TOXICITY FOR HUMAN PHARMACEUTICALS**

S5(R3)

Final version
Adopted on 18 February 2020



Default DART testing under ICHS5(R3)

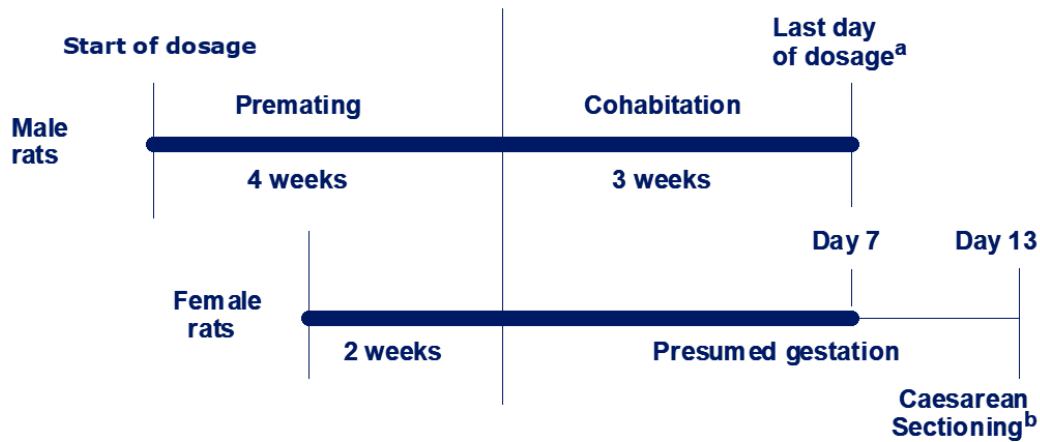
$\frac{C \ B \ G}{M \ E \ B}$

Effects of pharmaceuticals on reproductive cycle

Fertility and Early Embryonic Development (FEED)

Toxicity study

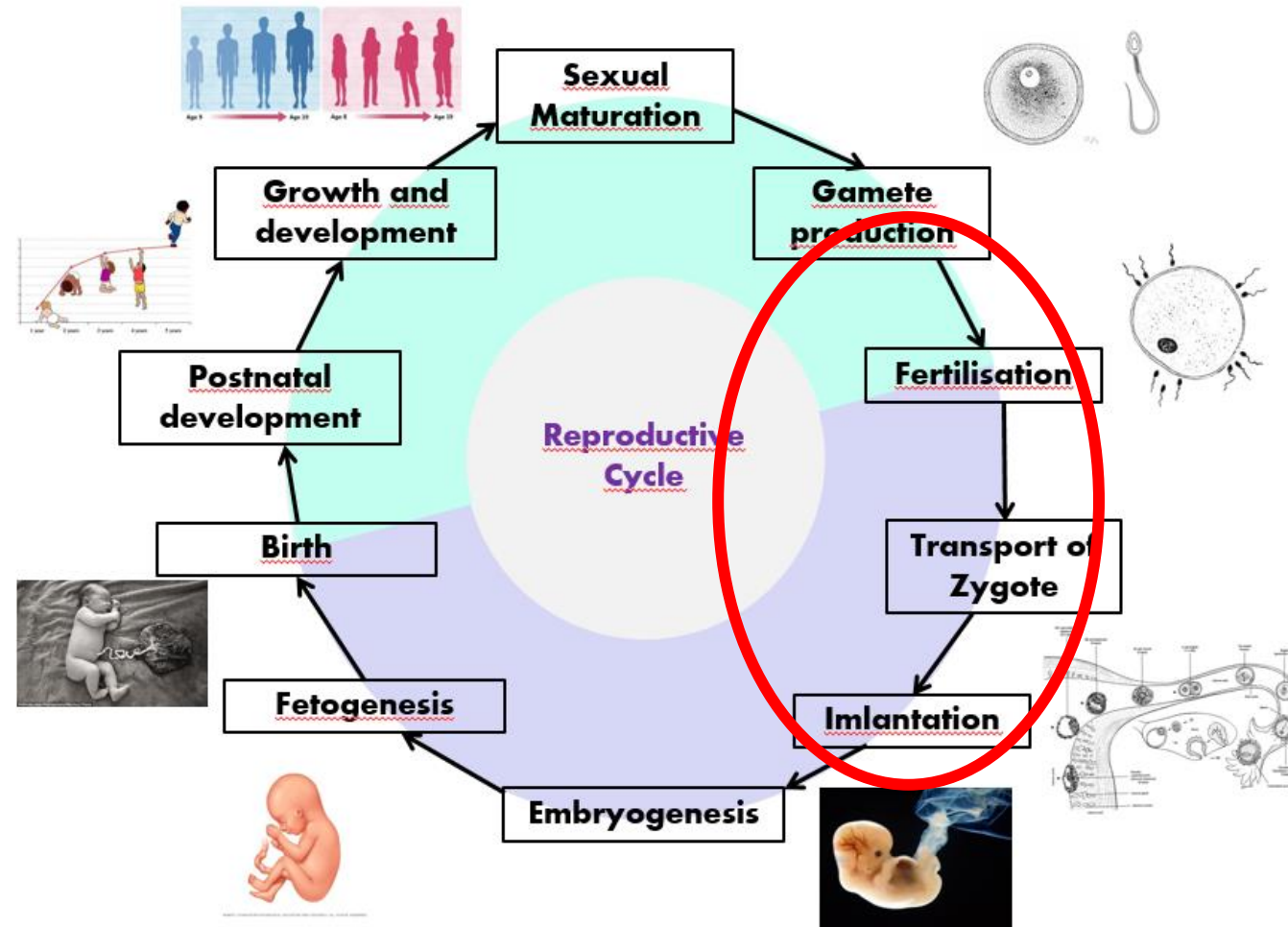
→ 2-4 weeks before conception – implantation



— = Dosage period

a = Male rats sacrificed, and sperm evaluations

b = Fetal evaluation (external only)



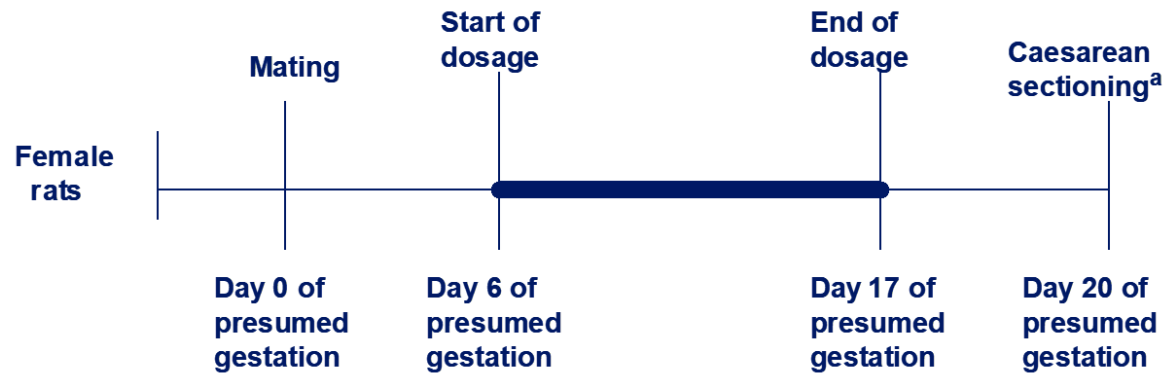
Default DART testing under ICHS5(R3)

$\frac{C \ B \ G}{M \ E \ B}$

Effects of pharmaceuticals on reproductive cycle

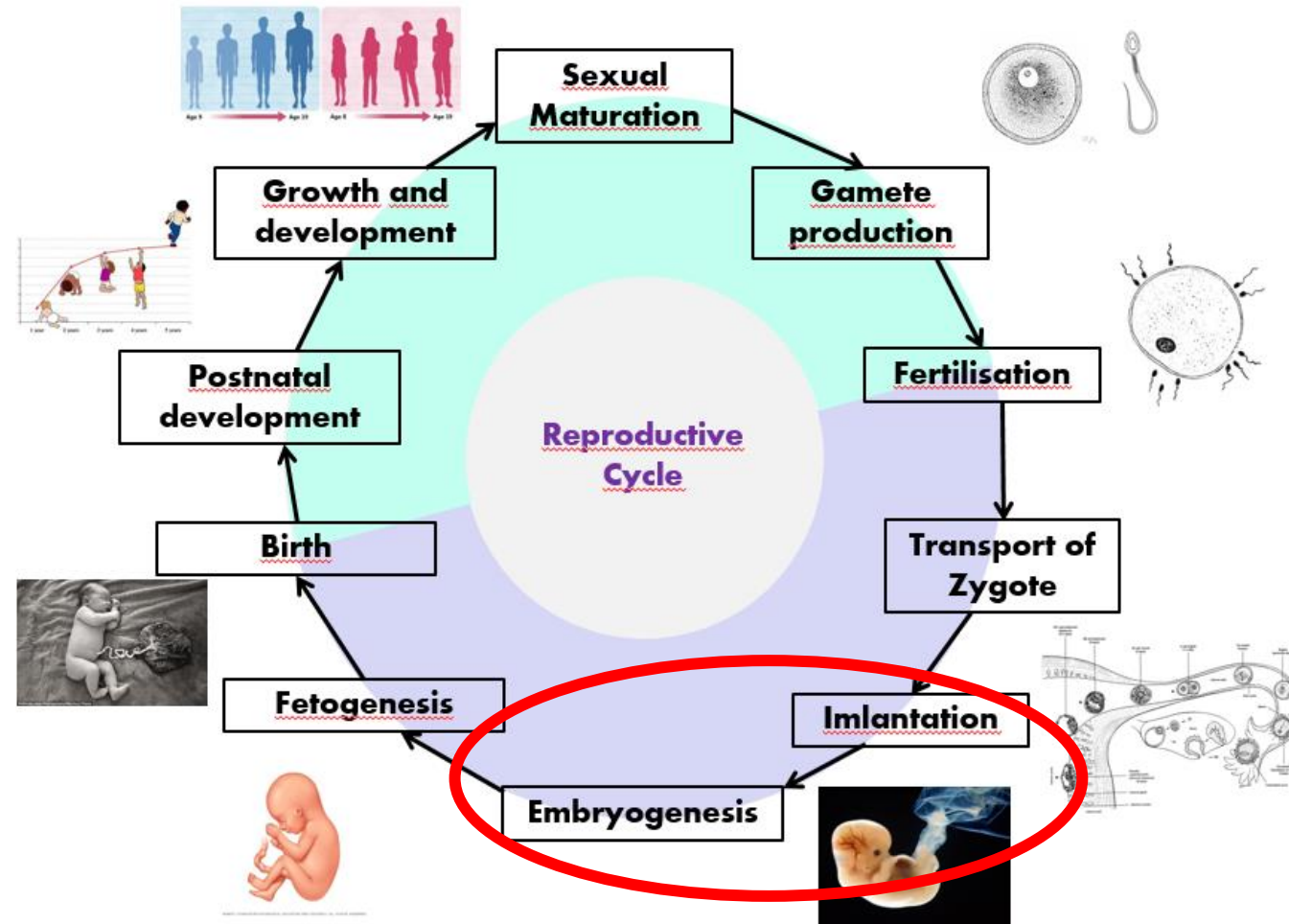
Embryo-fetal Developmental (EFD) Toxicity study

- Implantation – closure of hard palate
- Default 2 species (rodent and non-rodent)



— = Dosage period

a = Fetal evaluations (external, soft tissue and skeletal)



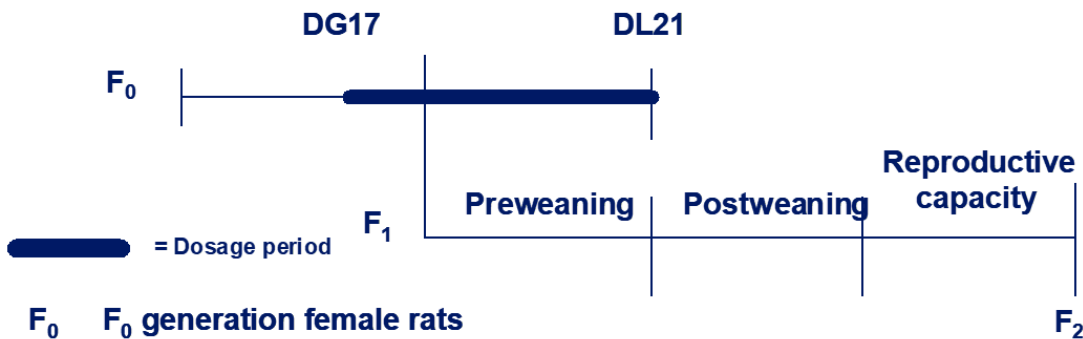
Default DART testing under ICHS5(R3)

$\frac{C \ B \ G}{M \ E \ B}$

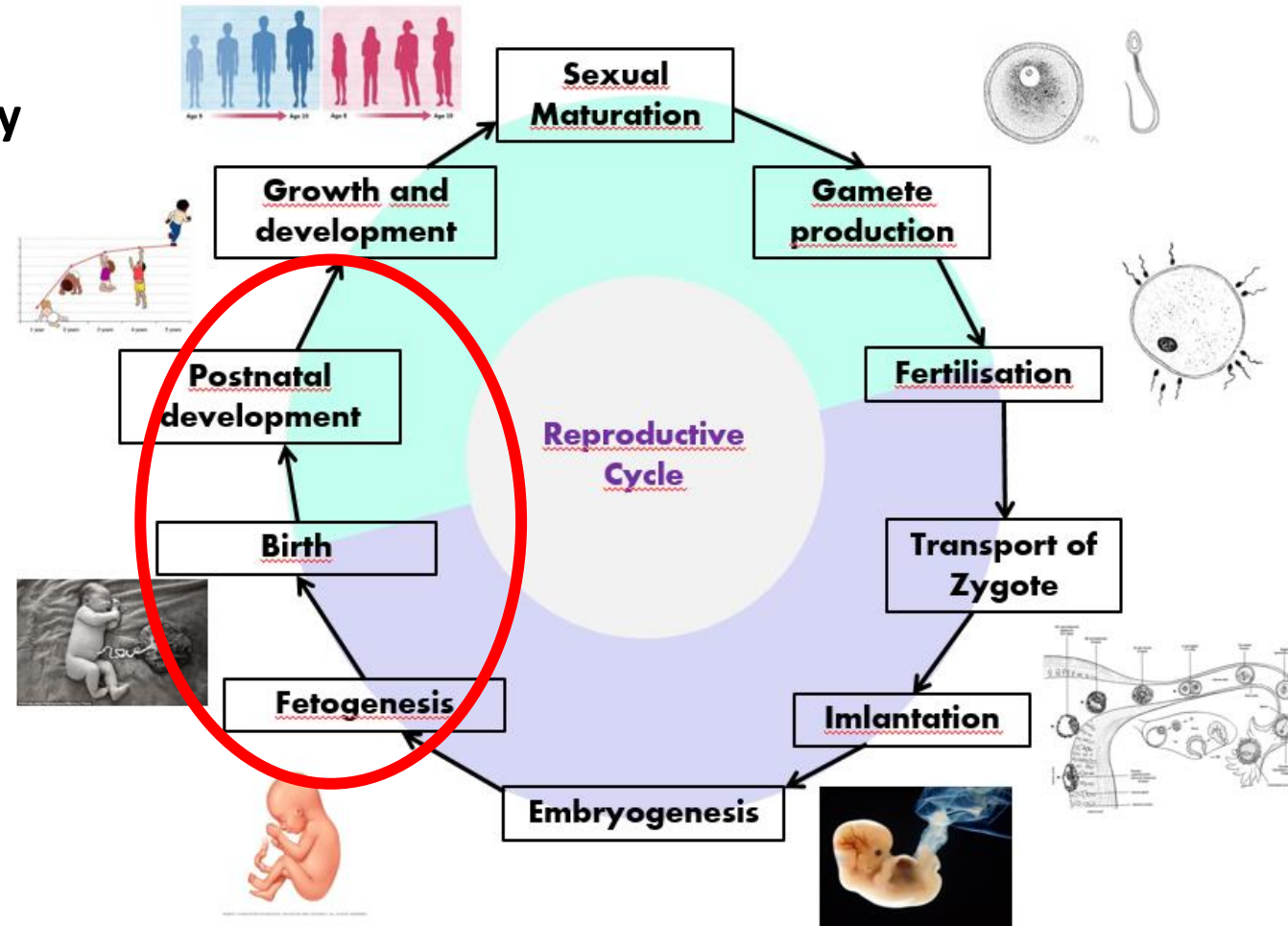
Effects of pharmaceuticals on reproductive cycle

Pre-post Natal Developmental (PPND) Toxicity study

- closure of hard palate - weaning
- Optional start at implantation
- Optional up to F1 generation



- F₀ F₀ generation female rats
- F₁ F₁ generation (offspring of F₀ generation)
- F₂ F₂ generation (offspring of F₁ generation)
- DG Day of gestation
- DL Day of lactation



New Approach Methods (NAMs) for EFD toxicity testing (history)

Classic models

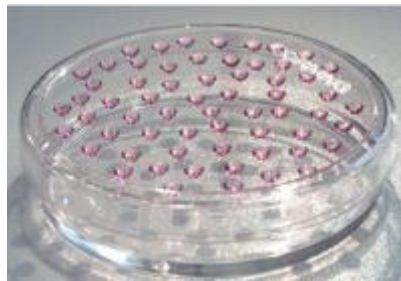
Whole Embryo Culture (WEC) (1970s)

Embryonic Stem Cell Test (EST) (1990s)

Zebrafish Embryo Toxicity Test (ZET) (2000s)

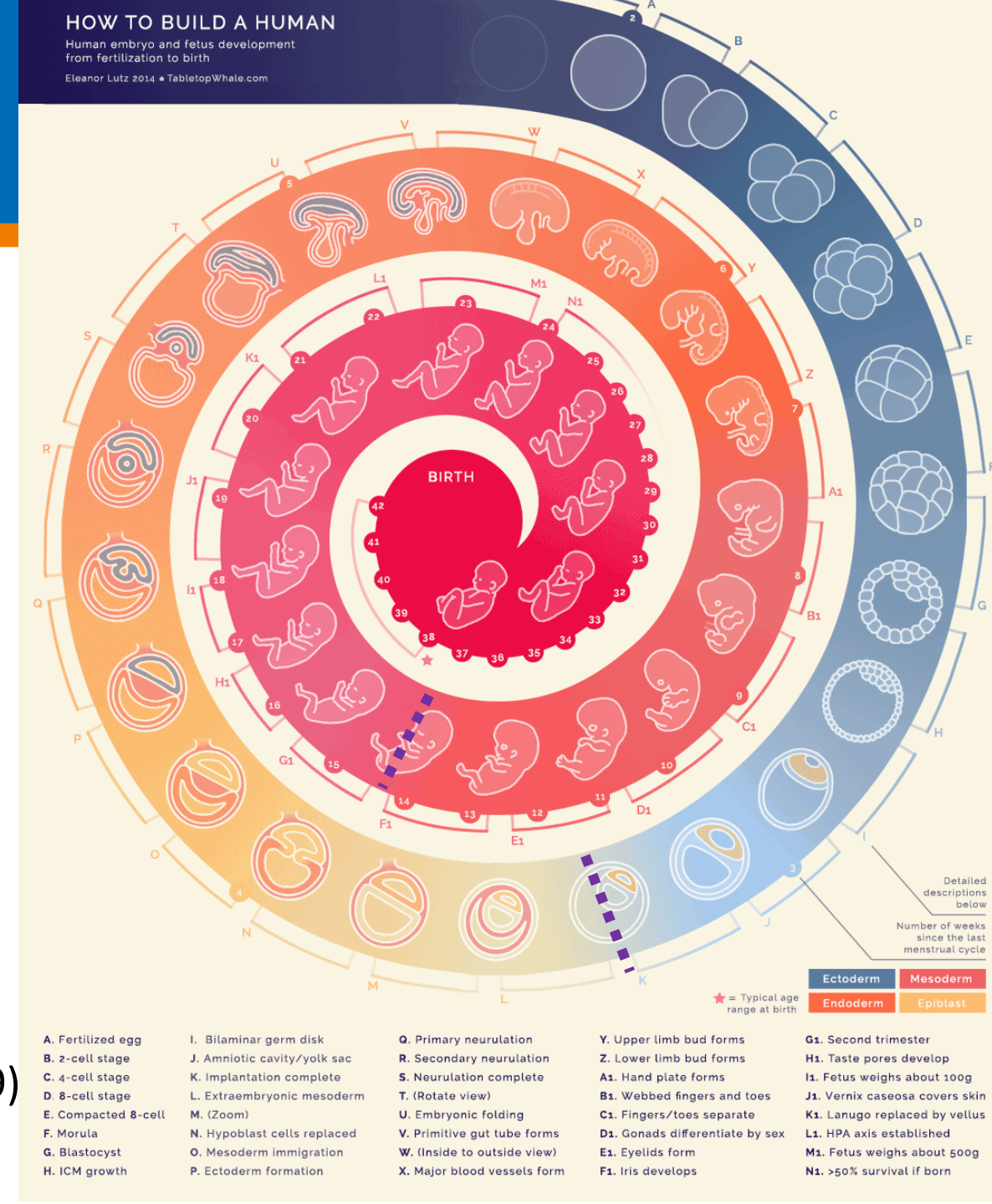


Aart Verhoef, RIVM



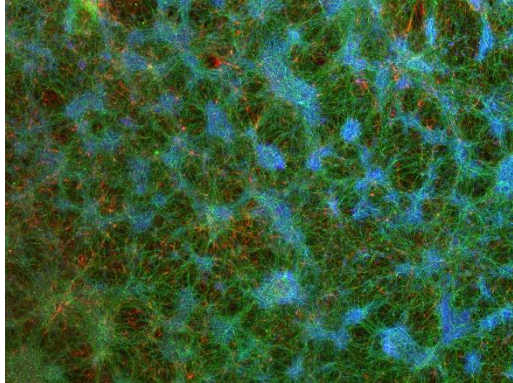
Sanne Hermsen, RIVM

- Investigate effects on development during window of implantation – closure hard palate
- Endpoints based on morphology
- Validation effort by ECVAM WEC/cardiac EST (2004-2009)



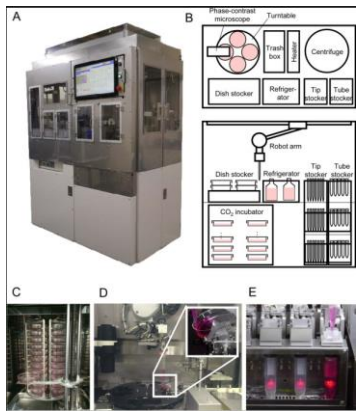
NAMs and innovation; Fast moving field!

Imaging



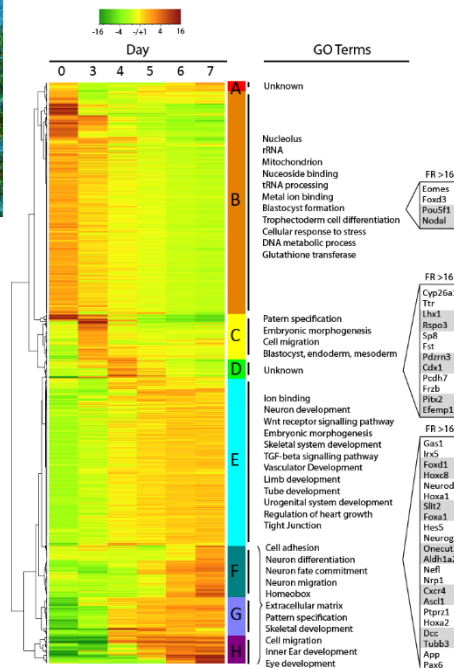
De Leuw, 2020

Robotics

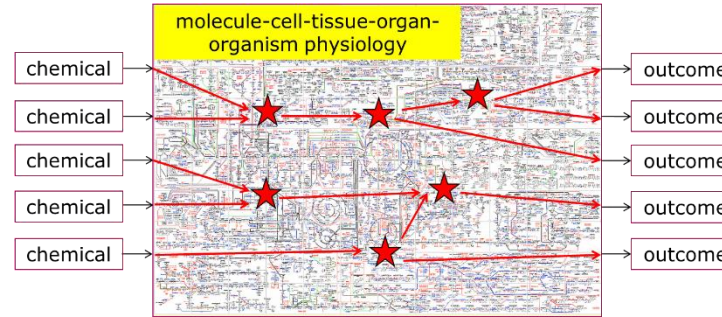


Konagaya, 2015

PCR / OMICS

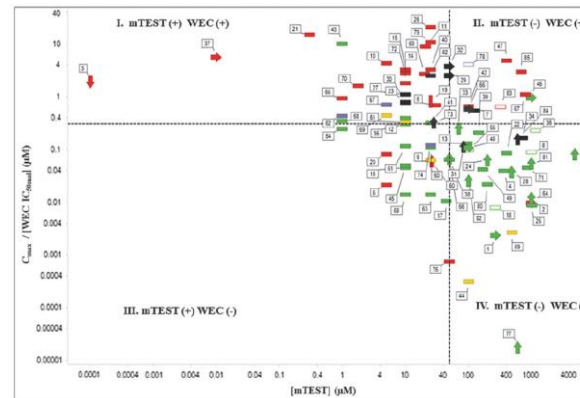


AOPs



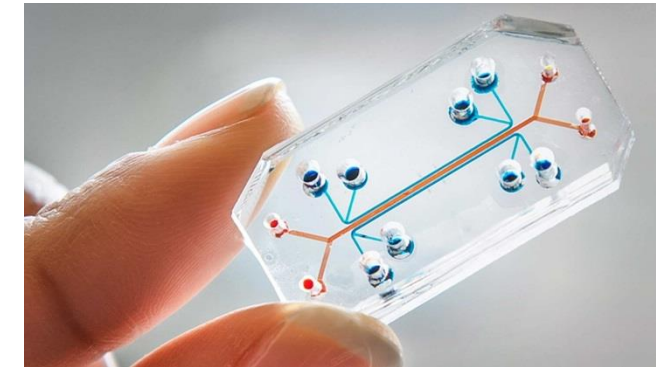
Aldert Piersma, RIVM

Tiered approach Testing Batteries

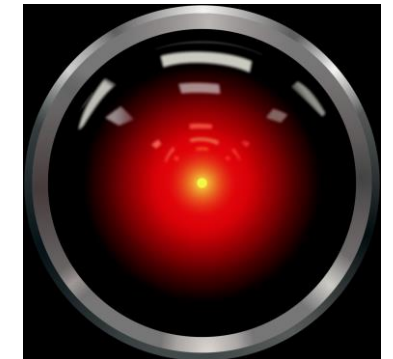


Green, 2018

Organ on Chip



Machine Learning / Artificial Intelligence



**ICH Topic S 5 (R2)
Detection of Toxicity to Reproduction for Medicinal Products & Toxicity to Male
Fertility**

Step 5

**NOTE FOR GUIDANCE ON THE DETECTION OF TOXICITY TO
REPRODUCTION FOR MEDICINAL PRODUCTS & TOXICITY TO MALE
FERTILITY
(CPMP/ICH/386/95)**

2.2. Other test systems

Other test systems are considered to be any developing mammalian and non-mammalian cell systems, tissues, organs, or organism cultures developing independently in vitro or in vivo. Integrated with whole animal studies either for priority selection within homologous series or as secondary investigations to elucidate mechanisms of action, these systems can provide invaluable information and, indirectly, reduce the numbers of animals used in experimentation. However, they lack the complexity of the developmental processes and the dynamic interchange between the maternal and the developing organisms. These systems cannot provide assurance of the **absence** of effect nor provide perspective in respect of risk/exposure. In short, there are no alternative test systems to whole animals currently available for reproduction toxicity testing with the aims set out in the introduction (Note 6).

NAMs under ICHS5(R3) (2020-present)

- 2010 Start of preparatory process at ICH level
- 2015 Official start of Revision procedure
- 2019 Step 4 approval by ICH
- 2020 Step 5 regional implementation

First ICH guidance to include information on use and qualification of NAMs as alternative for EFD testing

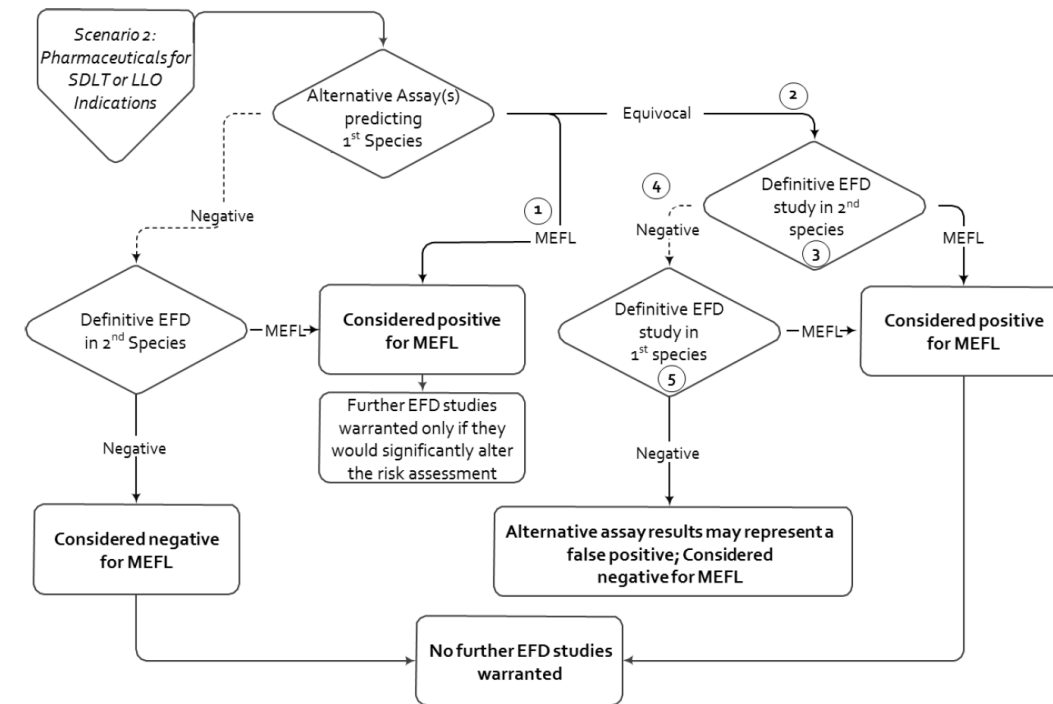
Because science develops quickly, and regulation does NOT... →

All information on NAMs and qualification in ANNEX → ICHS5(R4) maintenance procedure
- Possibility for 2 yearly changes to Annex

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When to USE NAMs under ICHS5(R3)

- To support Phase I + II clinical trials (=saving animals by attrition)
- **Qualified** alternative assays (predict **MEFL*** outcome in first species) + pEFD in a second species
- **Rodent** and **non-rodent** should be covered,
- Enable the limited inclusion of WOCBP (up to **150 WOCBP for up to 3 months**).
- Known **MoA** (class effects, known effect on developmental pathways) (ICHS5(R3)scheme figure 1 Annex 2, p39)
- No clinically relevant **exposure** possible in animals
- Support for WoE assessment when **equivocal** results in animal studies
- Indication for **severely debilitating** or **life-threatening diseases or late-life onset diseases**



ICHS5(R3) figure 2 Annex 2, p39

*MEFL = malformations and embryo-fetal lethality

Under ICHS5(R3), NAMs approaches should:

- provide a level of **confidence** for human safety assurance at least equivalent to that provided by the current testing paradigms.
- be **qualified** within a **certain context of use**, defined by
 - the **chemical applicability domain** of the assay, and
 - characterization of the **biological mechanisms** covered by the assay.

Qualification Criteria (ICHS5(R3), p36-37):

- Description and justification of predictive model
 - Which species does it predict Malformations and Embryo-fetal lethality (MEFL) for?
- Evaluation of biological plausibility of the model,
 - Mechanism of embryonic development in the model + adverse effects
 - Limitations of the models
 - Developmental window of prediction (in vivo)
- Determination of endpoints, and when negative or positive
- Statistical evidence to predict MEFL in a species (accuracy, prediction, sensitivity, specificity etc.)
- Historical data of the test system
- Reference compounds
 - list of training sets / test sets, source of data
 - Description of chemical domain predicted

Reference compound list for NAMs under ICHS5(R3)

29 example Reference Compounds are listed in Annex II and published by Andrews et al, 2019.



Analysis of exposure margins in developmental toxicity studies for detection of human teratogens

Paul A. Andrews^{a,*}, Diann Blanset^b, Priscila Lemos Costa^c, Martin Green^d, Maia L. Green^{e,1}, Abigail Jacobs^d, Rajkumar Kadaba^f, Jose A. Lebron^e, Britta Mattson^e, Mary Ellen McNERney^g, Daniel Minck^d, Luana de Castro Oliveira^c, Peter T. Theunissen^h, Joseph J. DeGeorge^{e,2}

^a Eisai Inc., Woodcliff Lake, NJ, USA
^b Boehringer Ingelheim Pharmaceuticals, Inc., Ridgefield, CT, USA
^c Agência Nacional de Vigilância Sanitária, Brasília, Brazil
^d US Food and Drug Administration, Silver Spring, MD, USA
^e Merck & Co, Inc., West Point, PA, USA
^f Health Canada, Ottawa, Ontario, Canada
^g Bristol-Myers Squibb, New Brunswick, NJ, USA
^h CBG-MEB, Utrecht, the Netherlands



Positive Controls	Human Teratogen	Rat MEFL	Rabbit MEFL
Acitretin	X	X	X
Aspirin	X	X	
Bosentan		X	
Busulfan	X	X	X
Carbamazepine	X	X	X
Cisplatin		X	
Cyclophosphamide	X	X	X
Cytarabine	X	X	
Dabrafenib		X	
Dasatinib		X	
Fluconazole	X	X	X
5-Fluorouracil	X	X	X
Hydroxyurea	X	X	X
Ibrutinib		X	X
Ibuprofen	X	X	
Imatinib		X	
Isotretinoin (13- <i>cis</i> -retinoic acid)	X	X	X
Methotrexate	X	X	X
Pazopanib		X	X
Phenytoin (Diphenylhydantoin)	X	X	X
Pomalidomide	presumed	X	X
Ribavirin		X	X
Tacrolimus		X	X
Thalidomide	X	X	X
Topiramate	X	X	X
Tretinoin (all- <i>trans</i> -retinoic acid)	X	X	X
Trimethadione	X	X	
Valproic acid	X	X	X
Vismodegib	presumed	X	

Cyclophosphamide

CAS No.: 50-18-0

B

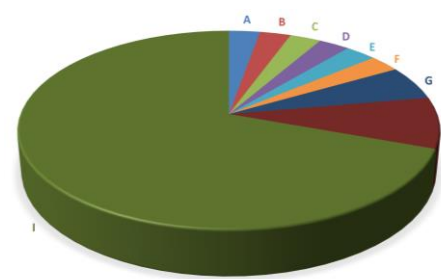
Rat NOAEL Dose C _{max} AUC	Rat LOAEL Dose C _{max} AUC	Rat Findings	Rabbit NOAEL Dose C _{max} AUC	Rabbit LOAEL Dose C _{max} AUC	Rabbit Findings	Human Dose C _{max} AUC	Margins NOAEL/Huma LOAEL/Human	Notes
NOAEL not identified (<2.5 mg/kg) [Chaube]	2.5 mg/kg IP GD9 [Chaube] <u>Cytoxan</u> C _{max} = 4.1 µg/mL ^a AUC = 3.65 µg·h/mL ^a <u>PM</u> C _{max} = 0.55 µg/mL ^b AUC _(0-24h) = 2.13 µg·h/mL ^b	<u>2.5 mg/kg GD9 [Chaube]</u> embryo-lethal <u>5 mg/kg GD11 [von Kreybig, Mirkes]</u> encephalocele, exencephaly, microcephaly, limb defects (ie, syndactyly and ectrodactyly), defective facial development (cleft palate)	NOAEL not identified (<30 mg/kg)	30 mg/kg IV single doses on GD6-14 [Mirkes, Fritz] <u>Cytoxan</u> C _{max} = 151 µg/mL ^c AUC _(0-8h) = 24.1 µg·h/mL ^d <u>PM</u> C _{max} = 0.07 µg/mL ^e AUC _(0-8h) = 0.297 µg·h/mL ^e	embryo-fetal resportions, omphalocele, cleft lip/palate, forelimb skeletal defects	1600 mg/m ² (40 mg/kg) IV (highest dose, q 3 - 4 weeks) ^f <u>Cytoxan</u> C _{max} = 106 µg/mL ^g AUC = 798 µg·h/mL ^g <u>PM</u> C _{max} = 14.4 µg/mL ^h AUC = 352 µg·h/mL ^h	NOAEL: <u>rat</u> : NOAEL not identified, but LOAEL margins were <0.1 <u>rabbit</u> NOAEL not identified, but LOAEL margins were <1.5 LOAEL: <u>rat</u> C _{max} : 0.04 (4.1/106) AUC: 0.005 (3.65/798)	<ul style="list-style-type: none"> MW CP = 261.086 MW PM = 221.018 Cytoxan is a prodrug, MEFL has been attributed to both phosphoramidate mustard (PM) and acrolein metabolites

Regulatory state of ICHS5(R3)

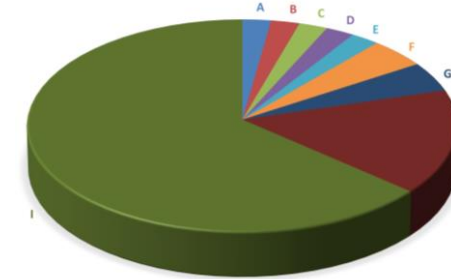
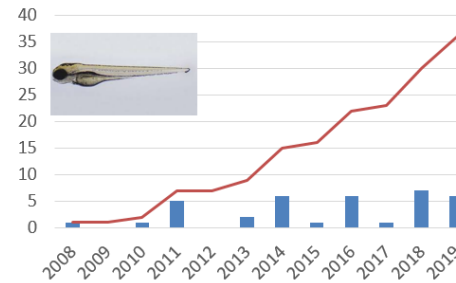
Since implementation of ICHS5(R3)

- No qualification exercises started at EMA
- One interested party at FDA, but not pursued further

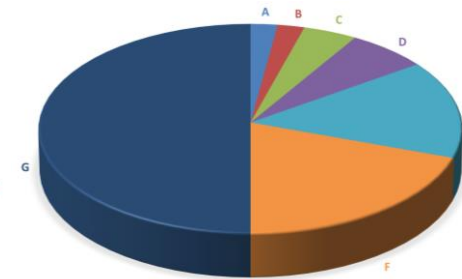
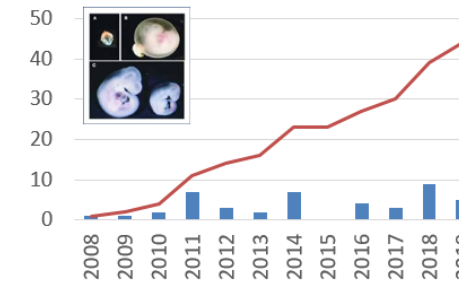
Companies do generate data in house
(60% of responders to IQ survey, not published)
Sometimes shared through submissions



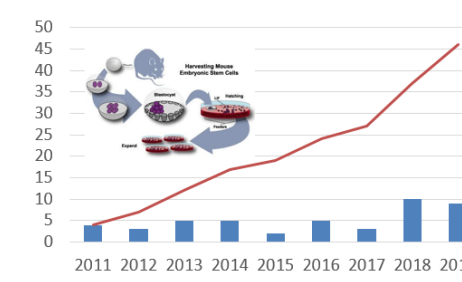
Zebrafish



Whole Embryo Culture



Embryonic Stem Cells



Companies are uncertain about qualification: what is expected, consequences
Regulators need to get more experienced with NAMs for regulatory purposes

IMPASSE...

Figures kindly provided by Ronald Wange, FDA

Breaking the Impasse and get Qualification moving at EMA

EMA wants to kickstart the Qualification of NAMs

3Rs Working Party restarted in 2022

- Updating and new **guidance** on qualification
- **Support** for qualification applications
- European Specialised Expert Community (**ESEC**) for NAMs
- Creation of a **worldwide cluster** of regulators for global alignment



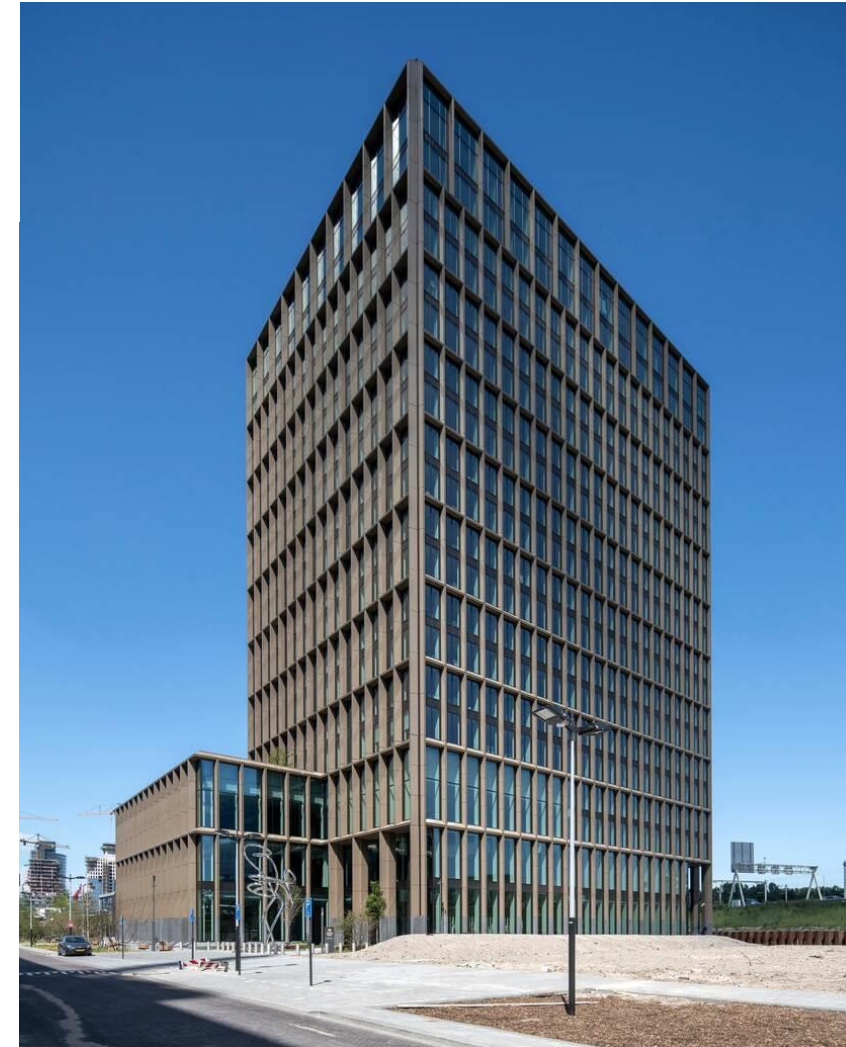
Innovative Task Force (ITF)

- **Informal** talks with EMA experts and ESEC members
- Discuss **proof of concept** and possibilities for qualification
- **Free** of charge

Formal talks with EMA before qualification (3RsWP / SAWP advice)

Formal Qualification (SAWP qualification advice)

→ EMA **certificate** that NAMs can be used under a certain context of use.



- Find common ground with industry to start qualifications →
- Increase in Qualification applications →
- Increased regulatory experience →
- More scenarios in which NAMs can be used under ICHS5(R3) Annex II



- Obtain more human relevant mechanistic data
- Decreased use of test animals
- Increased relevance for Labeling for Pregnant women, Lactation and Fertility



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